Terrestrial Animal Health Standards Commission Report September 2016

CHAPTER 12.10.

INFECTION WITH BURKHOLDERIA MALLEI (GLANDERS)

Article 12.10.1.

General provisions

Most glanders susceptible animals are equids. Equids are the major hosts and reservoirs of glanders although society can be are not available for on the occurrence of infection in zebras. Camelids and various carnivores including bears, canids and felids can also be infected but play no significant epidemiological role in the epidemiology of the disease. Glanders is a significant and potentially fatal zoonotic disease with fatal outcome if not treated in a timely manner.

For the purposes of the Terrestrial Code, glanders is defined as an infection of equids with Burkholderia mallei in an equid with or without the presence of clinical signs.

The chapter deals not only with the occurrence of clinical signs caused by *B. mallei*, but also with the presence of *infection* with *B. mallei* in the absence of clinical signs.

The following defines the occurrence of an infection with B. mallei.

- 1) B. mallei has been isolated from a sample from an equid; or
- 2) antigen or genetic material specific to B. mallei has been identified in a sample from an equid showing clinical or pathological signs consistent with glanders, or epidemiologically linked to a confirmed or suspected outbreak of glanders, or giving cause for suspicion of previous contact with B. mallei, or
- 3) antibodies specific to B. mallei have been identified by a testing regime appropriate to the species in a sample from an equid showing clinical or pathological signs consistent with glanders, or epidemiologically linked to a confirmed or suspected outbreak of glanders, or giving cause for suspicion of previous contact with B. mallei.

For the purposes of the *Terrestrial Code*, the *infective period* of *B. mallei* in equids is lifelong and the *incubation period* is six months.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 12.10.2.

Country or zone free from infection with B. mallei infection

A country or a zone that does not comply with the point 1 a) of Article 1.4.6. may be considered free from infection with B. mallei when:

- glanders infection with B. mallei is has been a notifiable disease in the entire country for at least the past three years;
- 2) either:
 - a) there has been no <u>case</u> outbreak and no evidence of infection with B. mallei in equids during the past three years following the destruction of the last case; or
- no evidence of infection with B. mallei has been found during the past six months following the destruction of the last case; and there is a surveillance programme in place demonstrating the absence of infection in accordance with Article 12.10.8. has demonstrated no evidence of infection with B. mallei in the past 42 six months;

AND

43) imports of equids and their germplasm into the country or zone are carried out in accordance with this chapter.

Article 12.10.3.

Recovery of free status

When a case is detected in a previously free country or zone, freedom from infection with B. mallei can be regained after the following:

- 1) a standstill of movements of equids and their germplasm from *establishments* affected infected or suspected of being affected infected has been imposed until the destruction of the last *case*;
- 2) an epidemiological investigation (trace-back, trace-forward), including investigations to determine the likely source of the *outbreak*, have has been carried out;
- 3) a *stamping-out policy*, which includes <u>at least</u> the destruction of all infected equids and cleansing and *disinfection* of the <u>affected infected</u> establishments, has been applied;
- 4) increased surveillance in accordance with Article 12.10.8. has been carried out and has <u>demonstrated</u> net detected any no evidence of infection in the six months after stamping-out disinfection of the last infected establishment and during that period measures have been in place to control the movement of equids.
- 5) measures are in place to control the movement of equids to prevent the spread of B. mallei.

When the measures above are not carried out, Article 12.10.2. applies.

Article 12.10.4.

Recommendations for importation of equids from countries or zones free from $\underline{infection}$ with B. mallei $\underline{infection}$

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the equid:

- 1) showed no clinical signs of glanders infection with B. mallei on the day of shipment;
- 2) either:
 - a) was kept for six months prior to shipment, or since birth, in <u>a</u> the exporting country or zone <u>free from infection</u> with <u>B. mallei</u>, or
 - b) was imported in accordance with Article 12.10.5., kept in an establishment in the exporting country for at least 30 days and then was subjected to a prescribed test with negative result on a sample taken during the 10 days prior to shipment.

Article 12.10.5.

Recommendations for importation of equids from countries or zones considered infected not free from infection with B. mallei

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the equid:

- 1) showed no clinical signs of glanders infection with B. mallei on the day of shipment;
- was kept for six months prior to shipment, or since birth, in an establishment where no case of glanders infection with B. mallei was reported during the six 12 months prior to shipment;

3) was <u>isolated and</u> subjected to <u>two</u> <u>a prescribed</u> test<u>s</u>, with negative result<u>s</u> on a sample<u>s</u> taken <u>during the</u> 30 days <u>apart with the second sample taken within 10 days</u> prior to shipment.

Article 12.10.6.

Recommendations for the importation of equine semen

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:

- 1) the donor males animals:
 - a) showed no clinical signs of glanders <u>infection with B. mallei</u> on the day of collection; and for the following 21 days;
 - b) were examined clinically for signs of orchitis, with negative results; were kept continuously:
 - i) either for a period of at least 21 days prior to, and for until at least 21 days after, the collection in a country or a zone free from infection with B. mallei, or
 - ii) for at least six months prior to the collection of the semen and during the collection in an establishment or artificial insemination centre free from infection with B. mallei and were subjected to a prescribed test, with a negative result on a sample taken between 21 and 30 days before the collection, or in the case of frozen semen between 21 and 30 days after the collection;
- 2) the semen was collected, processed and stored in accordance with the <u>relevant</u> recommendations in Chapter 4.5. <u>and in Articles 4.6.5. to 4.6.7.</u>

Article 12.10.7.

Recommendations for the importation of in vivo derived equine embryos

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:

- 1) the donor females animals:
 - a) showed no clinical signs of glanders infection with B. mallei on the day of collection and for the following 21 days;
 - b) were kept continuously:
 - i) either for a period of at least 21 days before, and for until at least 21 days after, the day of collection of the embryos in a country or a zone free from infection with B. mallei, or
 - ii) for at least six months prior to the collection and during the collection in an establishment free from infection with B. mallei and were subjected to a prescribed test, with a negative result on a sample taken between 21 and 30 days before the collection, or in the case of frozen embryos, between 21 and 30 days after the collection;
- 2) the embryos were collected, processed and stored in accordance with the <u>relevant</u> recommendations in Chapters 4.7. and 4.9., as relevant;
- the semen used for embryo production to fertilise the occytes complies with the recommendations in Article 12.10.6.

Article 12.10.8.

General Principles of surveillance

The purpose of surveillance is to determine the status of a country or a zone with respect to infection with B. mallei.

Surveillance should be carried out in accordance with Chapter 1.4.

Populations of captive wild, feral and wild equids should be included in the surveillance programme, for example through testing of roadkill or equids or culled as part of population control measures.

Clinical surveillance aims at detecting signs of glanders by close physical examination of susceptible animals. Clinical inspection is an important component of surveillance contributing to the desired level of confidence of detection of disease, if so long as a sufficiently large number of clinically susceptible animals is examined. Laboratory investigations should be conducted on all suspected cases.

Systematic pathological surveillance is an effective approach for glanders and should be conducted on dead equids on farm, at slaughterhouses/abattoirs and establishments for the disposal of carcasses of equids. Suspicious pathological findings should be confirmed by agent identification and isolates should be typed.

When conducting serological surveillance repeated testing of the equine population is necessary to reach an acceptable level of confidence.

Clinical examination and laboratory testing should be applied to clarify the status of suspects detected by either of these complementary diagnostic approaches. Laboratory testing and necropsy may contribute to confirm clinical suspicion, while clinical examination may contribute to confirmation of positive serology.

This article and Article 12.10.9, provide recommendations for surveillance for glanders and are complementary to Chapter 1.4. The impact and epidemiology of glanders vary in different regions of the world. The surveillance strategies employed for determining glanders status should be adapted to the respective epidemiological situation.

The surveillance programme should be designed to demonstrate that susceptible populations in a country or zone show no evidence of infection with B. mallei or to detect its introduction into a free population. If B. mallei is known to be present, surveillance should allow the estimation of the prevalence and the determination of the distribution of the infection.

A surveillance system in accordance with Chapter 1.4 should be under the responsibility of the Veterinary Authority and should have in place:

- a) a formal and ongoing system for detecting and investigating outbreaks of disease:
- <u>a procedure for the rapid collection and transport of samples from suspected cases to a laboratory with appropriate testing capability for glanders diagnosis;</u>
- c) a system for recording, managing and analysing diagnostic, epidemiological and surveillance data;
- d) established links with an OIE Reference Laboratory in case of need for confirmatory testing.

The glanders surveillance programme should include an early detection system for reporting suspected cases. Diagnosticians and those with regular contact with susceptible or infected equids should report promptly any suspicion of glanders to the Veterinary Authority. The reporting system under the Veterinary Authority should be supported directly or indirectly (e.g. through private veterinarians or veterinary paraprofessionals) by government awareness programmes. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in glanders, epidemiological evaluation and control as part of their contingency plan.

The Veterinary Authority should implement, when relevant, regular and frequent clinical inspections and random or targeted serological surveys and laboratory testing of high-risk groups or those adjacent to a country or zone infected with *B. mallei*. An effective surveillance system is likely to identify suspected cases that require follow-up investigation to confirm or exclude that the cause of the condition is *B. mallei*. All suspected cases of infection with *B. mallei* should be investigated immediately and samples should be taken and submitted to a laboratory. This requires that sampling kits and other equipment be available to those responsible for the surveillance. Details of the occurrence of suspected cases and how they were investigated and dealt with should be documented. This should include the results of diagnostic testing and the control measures to which the equids concerned were subjected during the investigation (quarantine, movement control).

Susceptible captive wild, feral and wild equine populations should be included in the surveillance programme.

Surveillance should address not only the occurrence of clinical signs caused by B. mallei, but also evidence of infection with B. mallei in the absence of clinical signs.

Article 12.10.9.

Surveillance strategies

The strategy employed may be based on clinical investigation, or randomised or targeted sampling at an acceptable level of statistical confidence. If glanders is present, it is usually at a very low prevalence. If an increased likelihood of infection in particular geographical locations or subpopulations can be identified, targeted sampling is appropriate.

To detect infection or to determine the distribution and estimate the prevalence of infection either at the level of the entire population or within targeted subpopulations, the design of the sampling strategy and frequency of testing should incorporate epidemiologically appropriate design prevalence for the selected populations. The sample size selected for testing should be statistically relevant to detect the presence of infection if it were to occur at a predetermined minimum rate. The design prevalence and confidence level should be consistent with the objectives of the surveillance and the epidemiological situation.

To substantiate freedom from infection in a country or zone, surveillance should be conducted in accordance with the relevant provisions of Chapter 1.4. Irrespective of the approach selected, the sensitivity and specificity of the diagnostic tests employed should be considered in the design and in the interpretation of the results obtained. The occurrence of false positive reactions has to be considered and the rate at which these false positives are likely to occur should be calculated in advance. Every positive result should be investigated to determine whether it is indicative of infection or not. This involves supplementary tests, trace-back and trace-forward, and inspection of individual animals and herds for clinical signs. Laboratory results should be interpreted in the context of the epidemiological situation.

Methods should include clinical surveillance and laboratory testing. They should always be applied in series to clarify the status of suspected cases of glanders detected by either of these complementary diagnostic approaches. Agent identification should be carried out on any equid positive or showing clinical signs. Any epidemiological unit within which suspected cases are detected should be considered infected until contrary evidence is produced.

Clinical surveillance

Clinical surveillance aims at detecting clinical signs by close physical examination of equids. However, clinical surveillance is of limited use only as asymptomatic carrier animals are the main reservoir of the disease.

2. Pathological and bacteriological surveillance

Systematic pathological surveillance is an effective approach for the detection of glanders and should be conducted on dead equids on farms, at slaughterhouses/abattoirs and facilities for the disposal of carcasses of equids. Suspicious pathological findings should be confirmed by agent identification and isolates should be characterised.

3. Serological surveillance

Serological surveillance for glanders is the preferred strategy. Repeated testing of the equid population with recommended tests is necessary to reach an acceptable level of confidence.

4. Malleinisation

<u>Frequently used as a surveillance method, malleinisation demonstrates hypersensitivity to antigens of B. mallei. However, this method has shortcomings that should be considered when interpreting results.</u>

Text deleted.